

### Era of Re-writing the (Livestock) Genome

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### **Genetic Engineering (of livestock):**

### Transgeneis (GM) - since 1985 Genome Editing - rewrite the genome





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# Can we use these tools differently? >> yes and no









#### No – both allow gene addition / deletion

### So - what can we do?

- accelerate genetics
- by-pass genetics









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#### Livestock Breeding: we are good at it



### **Accelerate Genetics:**

increasing fixation rate for low frequency variation

eliminating deleterious alleles

Pawnee Farm Arlinda Chief

recombinetics



New edits to animal genes cut down on ro

multiple SNPs at the same time

**Standard selection traits**: reproductive performance,



maternal ability, growth rate, feed efficiency, longevity, carcass merit / milk production



# ROSLN

#### **Promotion of Alleles by Genome Editing (PAGE)**

- weakness of GS with perfect accuracy is that alleles do not segregate independently
- with PAGE alleles behave as though they segregate independently (offers precision)
- genomic selection decoupled selection from phenotyping ... genome editing decouples gain from selection





#### **Promotion of Alleles by Genome Editing (PAGE)**







#### Livestock Breeding: use what already there







### **By-pass genetics:**

- introduce novel variation (e.g. Nanos2, CD163)
- "capturing" rare breed alleles or from different species (e.g. RELA)
- disease 'resistance'
- single sex offspring
- adaption to stress (e.g. temperature)
- welfare traits







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#### Gene-edited pigs are protected from porcine reproductive and respiratory syndrome virus

Kristin M Whitworth, Raymond R R Rowland, Catherine L Ewen, Benjamin R Trible, Maureen A Kerrigan, Ada G Cino-Ozuna, Melissa S Samuel, Jonathan E Lightner, David G McLaren, Alan J Mileham, Kevin D Wells & Randall S Prather

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Bacteria binding ASFV binding putative TWEAK binding sites < HbHp binding PRRSV GP2/GP5 uncoating interaction PRRSV-binding in soluble form putative TWEAK SRCR domain binding sites PST segment cell membrane cytoplasmic tail PRRSV replication efficiency

#### RESEARCHARTICLE

PLOS PATHOGENS

Precision engineering for PRRSV resistance in pigs: Macrophages from genome edited pigs lacking CD163 SRCR5 domain are fully resistant to both PRRSV genotypes while maintaining biological function

Christine Burkard<sup>1</sup>, Simon G. Lillico<sup>1</sup>, Elizabeth Reid<sup>2</sup>, Ben Jackson<sup>2</sup>, Alan J. Mileham<sup>3</sup>, Tahar Ait-Ali<sup>1</sup>, C. Bruce A. Whitelaw<sup>1</sup>, Alan L. Archibald<sup>1</sup>

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#### SCIENTIFIC REPORTS

#### OPEN Mammalian interspecies substitution of immune modulatory alleles by genome editing

Received: 20 October 2015 Accepted: 27 January 2016 Published: 22 February 2016

<sup>5</sup> Simon G. Lillico<sup>1</sup>, Chris Proudfoot<sup>1</sup>, Tim J. King<sup>1</sup>, Wenfang Tan<sup>1</sup>, Lei Zhang<sup>3</sup>, Rachel Mardjuki<sup>2</sup>, David E. Paschon<sup>1</sup>, Edward J. Rebar<sup>2</sup>, Fyodor D. Urnov<sup>2</sup>, Alan J. Mileham<sup>3</sup>, David G. McLaren<sup>3</sup> 8. C. Bruce A. Whitelaw<sup>1</sup>

We describe a fundamentally novel feat of animal genetic engineering: the precise and efficient substitution of an agronomic haplotype into a domesticated species. Zinc finger nuclease in-embryo editing of the RELA locus generated live born domestic pigs with the warthog RELA orthologue, associated with resilience to African Swine Fever. The ability to efficiently achieve interspecies allele introgension in one generation opens unprecedented opportunities for agriculture and basic research.











## SCIENTIFIC REPORTS

Received: 21 October 2016 Accepted: 02 December 2016 Published: 10 January 2017

#### **OPEN** Generation of germline ablated male pigs by CRISPR/Cas9 editing of the NANOS2 gene

Ki-Eun Park<sup>1,2,3,\*</sup>, Amy V. Kaucher<sup>4,\*</sup>, Anne Powell<sup>2</sup>, Muhammad Salman Waqas<sup>4</sup>, Shelley E.S. Sandmaier<sup>1,2</sup>, Melissa J. Oatley<sup>4</sup>, Chi-Hun Park<sup>1,2</sup>, Ahmed Tibary<sup>4</sup>, David M. Donovan<sup>2</sup>, Le Ann Blomberg<sup>2</sup>, Simon G. Lillico<sup>5</sup>, C. Bruce A. Whitelaw<sup>5</sup>, Alan Mileham<sup>6</sup>, Bhanu P. Telugu<sup>1,2,3</sup> & Jon M. Oatley<sup>4</sup>

Genome editing tools have revolutionized the generation of genetically modified animals including livestock. In particular, the domestic pig is a proven model of human physiology and an agriculturally important species. In this study, we utilized the CRISPR/Cas9 system to edit the NANOS2 gene in pig embryos to generate offspring with mono-allelic and bi-allelic mutations. We found that NANOS2 knockout pigs phenocopy knockout mice with male specific germline ablation but other aspects of testicular development are normal. Moreover, male pigs with one intact NANOS2 allele and female knockout pigs are fertile. From an agriculture perspective, NANOS2 knockout male pigs are expected to serve as an ideal surrogate for transplantation of donor spermatogonial stem cells to expand the availability of gametes from genetically desirable sires.







Likely next steps for genome editing in livestock:

Short term = focus on disease traits

Short to medium term = surrogate sires

Medium term = fix up genetic load and deleterious mutations

Long term = PAGE for quantitative traits

























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